

STEREOCHEMISTRY ABSTRACTS

M. Yokoyama, T. Sugai, and H. Ohta

Tetrahedron: Asymmetry 1993, 4, 1081



(*S*)- α -Methylnorleucine hydrochloride

E.e. = >99% (by chiral HPLC of *N*-Cbz-*O*-Me deriv.)

$[\alpha]^{23}_D +10$ (c 2.0, 5*N* HCl)

Source of chirality: Methyl (*R*)-2-carbamoyl-2-methylhexanoate

Absolute configuration: 2*S*

(assigned by optical rotatory dispersion)

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Tetrahedron: Asymmetry 1993, 4, 1081



(*R*)-2-Cyano-2-methylhexanoic acid

E.e. = 96% (by chiral HPLC of β -naphthyl ester)

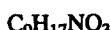
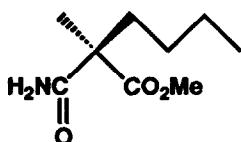
$[\alpha]^{19}_D +8.6$ (c 2.0, MeOH)

Source of chirality: Methyl (*R*)-2-carbamoyl-2-methylhexanoate

Absolute configuration: 2*R*

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Tetrahedron: Asymmetry 1993, 4, 1081



Methyl (*R*)-2-carbamoyl-2-methylhexanoate

E.e. = 96%

$[\alpha]^{20}_D -15.6$ (c 1.0, CHCl₃)

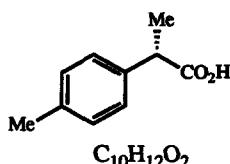
Source of chirality: enzymatic hydrolysis of prochiral dinitrile

Absolute configuration: 2*R*

(assigned via chemical correlation)

T. Beard, M.A. Cohen, J.S. Parratt, N.J. Turner,
J. Crosby, and J. Moilliet

Tetrahedron: Asymmetry 1993, 4, 1085



(*S*)-2-(4'-methylphenyl)-propionic acid

E.e. = >95%

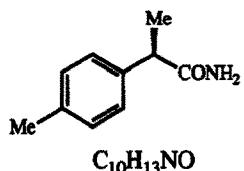
$[\alpha]_D^{25} = +57.0$ (c = 1.0, CHCl₃)

Source of chirality: enzyme nitrile hydrolysis

Absolute configuration: 2*S*

T. Beard, M.A. Cohen, J.S. Parratt, N.J. Turner,
J. Crosby, and J. Moilliet

Tetrahedron: Asymmetry 1993, 4, 1085



E.e. = >95%

$[\alpha]_D^{25} = -49.7$ ($c = 1.14, \text{CHCl}_3$)

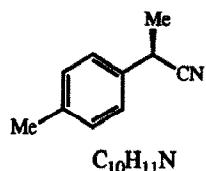
Source of chirality: enzyme nitrile hydrolysis

Absolute configuration: 2*R*

(*R*)-2-(4'-methylphenyl)-propionamide

T. Beard, M.A. Cohen, J.S. Parratt, N.J. Turner,
J. Crosby, and J. Moilliet

Tetrahedron: Asymmetry 1993, 4, 1085



E.e. = >95%

$[\alpha]_D^{25} = +13.1$ ($c = 1.1, \text{CHCl}_3$)

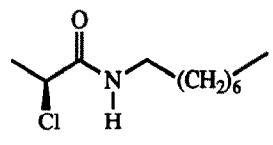
Source of chirality: enzyme nitrile hydrolysis

Absolute configuration: 2*R*

(*R*)-2-(4'-methylphenyl)-propionitrile

M. Quirós, V. M. Sánchez, R. Brieva, F. Rebolledo and V. Gotor

Tetrahedron: Asymmetry 1993, 4, 1105



E.e. 70% [by 1H-NMR spectroscopy using the chiral shift reagent tris[3-(trifluoromethylhydroxymethylene)-(+)-camphorato]europium (III).]

$[\alpha]_D^{25} = -10.0$ ($c 0.97, \text{CHCl}_3$)

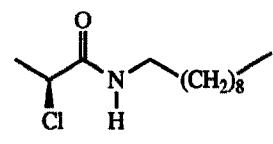
Source of chirality: Enzymatic aminolysis

Absolute configuration: S

(*S*)-2-Chloro-N-octylpropanamide

M. Quirós, V. M. Sánchez, R. Brieva, F. Rebolledo and V. Gotor

Tetrahedron: Asymmetry 1993, 4, 1105



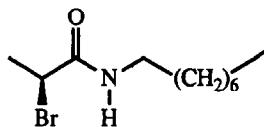
E.e. 92% [by 1H-NMR spectroscopy using the chiral shift reagent tris[3-(trifluoromethylhydroxymethylene)-(+)-camphorato]europium (III).]

$[\alpha]_D^{25} = -9.9$ ($c 1.02, \text{CHCl}_3$)

Source of chirality: Enzymatic aminolysis

Absolute configuration: S

(*S*)-2-Chloro-N-decylpropanamide

 $C_{11}H_{22}BrNO$

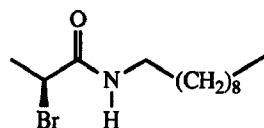
(S)-2-Bromo-N-octylpropanamide

E.e. 61% [by 1H-NMR spectroscopy using the chiral shift reagent tris[3-(trifluoromethylhydroxymethylene)-(+) -camphorato]europium (III).]

 $[\alpha]_D^{25} = -6.0$ (c 0.72, $CHCl_3$)

Source of chirality: Enzymatic aminolysis

Absolute configuration: S

 $C_{13}H_{26}BrNO$

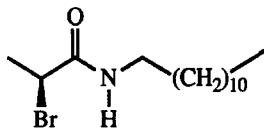
(S)-2-Bromo-N-decylpropanamide

E.e. 64% [by 1H-NMR spectroscopy using the chiral shift reagent tris[3-(trifluoromethylhydroxymethylene)-(+) -camphorato]europium (III).]

 $[\alpha]_D^{25} = -6.5$ (c 0.65, $CHCl_3$)

Source of chirality: Enzymatic aminolysis

Absolute configuration: S

 $C_{15}H_{30}BrNO$

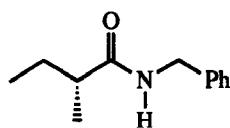
(S)-2-Bromo-N-dodecylpropanamide

E.e. 64% [by 1H-NMR spectroscopy using the chiral shift reagent tris[3-(trifluoromethylhydroxymethylene)-(+) -camphorato]europium (III).]

 $[\alpha]_D^{25} = -3.6$ (c 0.60, $CHCl_3$)

Source of chirality: Enzymatic aminolysis

Absolute configuration: S

 $C_{12}H_{17}NO$

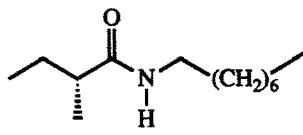
(R)-N-Benzyl-2-methylbutanamide

E.e. 78% [by comparison with an authentic sample obtained from (S)-(+)2-methylbutyric anhydride]

 $[\alpha]_D^{25} = -6.8$ (c 1.05, $CHCl_3$)

Source of chirality: Enzymatic aminolysis

Absolute configuration: R



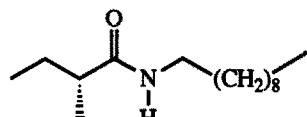
(R)-2-Methyl-N-octylbutanamide

E.e. 50% [by comparison with an authentical sample obtained from (S)-(+) -2-methylbutyric anhydride]

$[\alpha]_D^{25} = -6.0$ (c 0.99, CHCl_3)

Source of chirality: Enzymatic aminolysis

Absolute configuration: R



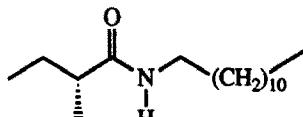
(R)-N-Decyl-2-methylbutanamide

E.e. 50% [by comparison with an authentical sample obtained from (S)-(+) -2-methylbutyric anhydride]

$[\alpha]_D^{25} = -3.7$ (c 0.96, CHCl_3)

Source of chirality: Enzymatic aminolysis

Absolute configuration: R



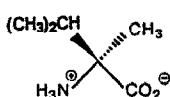
(R)-N-Dodecyl-2-methylbutanamide

E.e. 48% [by comparison with an authentical sample obtained from (S)-(+) -2-methylbutyric anhydride]

$[\alpha]_D^{25} = -4.6$ (c 0.70, CHCl_3)

Source of chirality: Enzymatic aminolysis

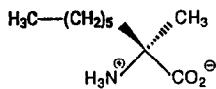
Absolute configuration: R

 $\text{C}_6\text{H}_{13}\text{NO}_2$
2-Amino-2,3-dimethylbutanoic acid

E.e.>98% [by NMR using S-2-chloropropionyl chloride]

$[\alpha]_D^{20} = -4.0$ (c 1.3, H_2O) HCl-salt, S-enantiomer

Source of chirality: enzymatic resolution



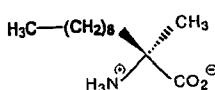
E.e.>99% [by HPLC]

$[\alpha]_D^{20} = +13.3$ (c1,1N HCl), S-enantiomer

Source of chirality: enzymatic resolution



2-Amino-2-methyloctanoic acid



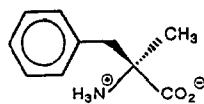
E.e.>99% [by HPLC]

$[\alpha]_D^{20} = +16.4$ (c0.5,MeOH) HCl-salt, S-enantiomer

Source of chirality: enzymatic resolution



2-Amino-2-methylundecanoic acid



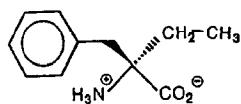
E.e.>99% [by HPLC]

$[\alpha]_D^{20} = -22.0$ (c1,H₂O), HCl-salt, S-enantiomer

Source of chirality: enzymatic resolution



2-Amino-2-methyl-3-phenylpropanoic acid



E.e.>96% [by HPLC]

$[\alpha]_D^{20} = -22.8$ (c1,H₂O), HCl-salt, S-enantiomer

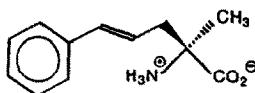
Source of chirality: enzymatic resolution



2-Amino-2-ethyl-3-phenylpropanoic acid

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



E.e.>98% [by HPLC]

$[\alpha]_D^{25}=-1.8$ (c1,1N HCl), S-enantiomer

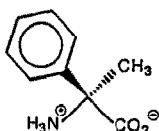
Source of chirality: enzymatic resolution

C₁₂H₁₅NO₂

2-Amino-2-methyl-5-phenylpent-4-enoic acid

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



E.e.=95% [by HPLC]

$[\alpha]_D^{20}=+86.0$ (c1,1N HCl), S-enantiomer

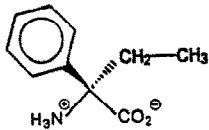
Source of chirality: enzymatic resolution

C₉H₁₁NO₂

2-Amino-2-methylphenylacetic acid

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



E.e.=94% [by HPLC]

$[\alpha]_D^{20}=+37.6$ (c1,1N HCl), S-enantiomer

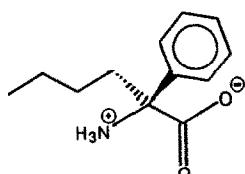
Source of chirality: enzymatic resolution

C₁₀H₁₃NO₂

2-Amino-2-ethylphenylacetic acid

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



E.e.=93% [by HPLC]

$[\alpha]_D^{20}=+25.3$ (c1,1N HCl) S(?)-enantiomer

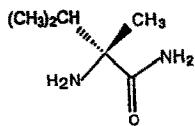
Source of chirality: enzymatic resolution

C₁₂H₁₇NO₂

2-Amino-2-phenylhexanoic acid

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



E.e.>98% [by NMR using S-2-chloropropionyl chloride]

$[\alpha]_D^{20}=+27.5$ (c1,H₂O) R-enantiomer

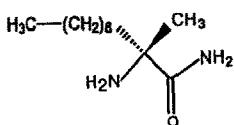
Source of chirality: enzymatic resolution



2-Amino-2,3-dimethylbutanoic acid amide

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



E.e.=99% [by HPLC]

$[\alpha]_D^{20}=-12.6$ (c1,1N HCl), R-enantiomer

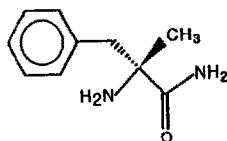
Source of chirality: enzymatic resolution



2-Amino-2-methylundecanoic acid amide

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



E.e.=99% [by HPLC]

$[\alpha]_D^{20}=+42.0$ (c1,MeOH), R-enantiomer

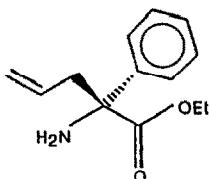
Source of chirality: enzymatic resolution



2-Amino-2-methyl-3-phenylpropanoic acid amide

B. Kaptein, W.H.J. Boesten, Q.B. Broxterman, P.J.H. Peters,
H.E. Schoemaker and J. Kamphuis.

Tetrahedron: Asymmetry 1993, 4, 1113



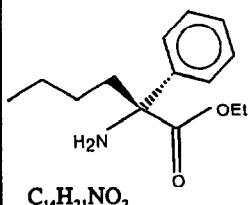
E.e.=95% [by HPLC]

$[\alpha]_D^{20}=-7.8$ (c1,1N HCl), R-enantiomer

Source of chirality: enzymatic resolution



Ethyl 2-amino-2-phenylpent-4-enoate



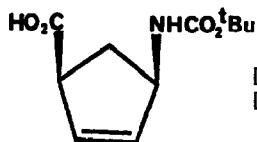
Ethyl 2-amino-2-phenylhexanoate

E.e.=97% [by HPLC]

$[\alpha]_D^{20} = -7.7$ (c1,1N HCl), R(?)-enantiomer

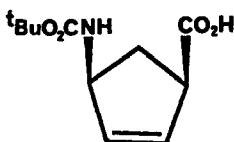
Source of chirality: enzymatic resolution

S. J. C. Taylor, R. McCague, R. Wisdom, C. Lee, K. Dickson, G. Ruecroft, F. O'Brien,
J. Littlechild, J. Bevan, S. M. Roberts and C. T. Evans



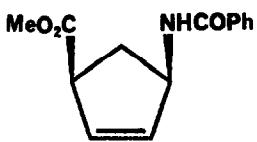
[1(R), 4(S)]-N-(tert-Butoxycarbonyl)-4-amino-2-cyclopentene-1-carboxylic acid
 $[\alpha]_D^{25} + 40.3$ (c = 2.1, CH₂Cl₂)

S. J. C. Taylor, R. McCague, R. Wisdom, C. Lee, K. Dickson, G. Ruecroft, F. O'Brien,
J. Littlechild, J. Bevan, S. M. Roberts and C. T. Evans



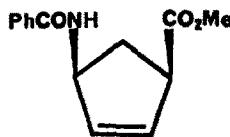
[1(S), 4(R)]-N-(tert-Butoxycarbonyl)-4-amino-2-cyclopentene-1-carboxylic acid
 $[\alpha]_D^{25} - 40.3$ (c = 2.1, CH₂Cl₂)

S. J. C. Taylor, R. McCague, R. Wisdom, C. Lee, K. Dickson, G. Ruecroft, F. O'Brien,
J. Littlechild, J. Bevan, S. M. Roberts and C. T. Evans



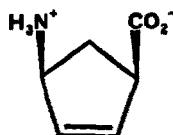
[1(R), 4(S)]-4-benzoylamino-2-cyclopentene-1-carboxylic acid, methyl ester
m.p. 80.5-82°C
 $[\alpha]_D^{25} - 33.6$ (c = 2.2, CH₂Cl₂)

S. J. C. Taylor, R. McCague, R. Wisdom, C. Lee, K. Dickson, G. Ruecroft, F. O'Brien,
J. Littlechild, J. Bevan, S. M. Roberts and C. T. Evans



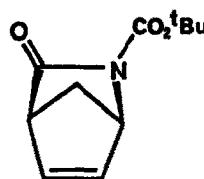
[1(S), 4(R)]-4-benzoylamino-2-cyclopentene-1-carboxylic acid, methyl ester
m.p. 80.5-82°C
[α]_D²⁵ + 33.6 (c = 2.2, CH₂Cl₂)

S. J. C. Taylor, R. McCague, R. Wisdom, C. Lee, K. Dickson, G. Ruecroft, F. O'Brien,
J. Littlechild, J. Bevan, S. M. Roberts and C. T. Evans



[1(R), 4(S)]-4-Amino-2-cyclopentene-1-carboxylic acid
[α]_D + 242 (c = 2, H₂O)

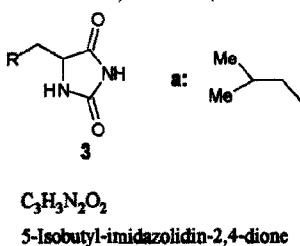
S. J. C. Taylor, R. McCague, R. Wisdom, C. Lee, K. Dickson, G. Ruecroft, F. O'Brien,
J. Littlechild, J. Bevan, S. M. Roberts and C. T. Evans



[1(R), 4(S)]-N-(tert-Butoxycarbonyl)-2-azabicyclo[2.2.1]hept-5-en-3-one
[α]_D - 189 (c = 0.89, CH₂Cl₂)

Enantioseparation of 5-Monosubstituted Hydantoins by Capillary Gas Chromatography - Investigation of Chemical and Enzymatic Racemization

H. Lickefett¹, K. Krohn², W.A. König³, B. Gehrcke³ and C. Syldatk¹



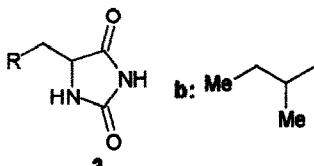
E. e > 98 %

Source of chirality : Chemical synthesis from the corresponding D- or L- or amino acid
Absolute configuration : R or S

Enantioseparation of 5-Monosubstituted Hydantoins by Capillary Gas Chromatography - Investigation of Chemical and Enzymatic Racemization

Tetrahedron: Asymmetry 1993, 4, 1129

H. Lickefett¹, K. Krohn², W.A. König³, B. Gehrcke³ and C. Syldatk¹



C₅H₈N₂O₂
5-(1-Methyl-propyl)-imidazolidin-2,4-dione

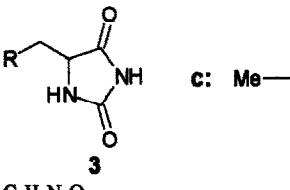
E. e > 98 %

Source of chirality : Chemical synthesis from the corresponding D- or L- α - amino acid
Absolute configuration : 2R, 3R or 2S, 3S

Enantioseparation of 5-Monosubstituted Hydantoins by Capillary Gas Chromatography - Investigation of Chemical and Enzymatic Racemization

Tetrahedron: Asymmetry 1993, 4, 1129

H. Lickefett¹, K. Krohn², W.A. König³, B. Gehrcke³ and C. Syldatk¹



C₄H₆N₂O₂
5-methyl-imidazolidin-2,4-dione

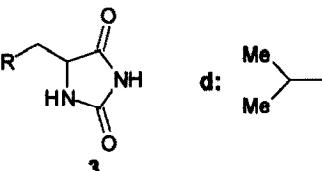
E. e > 98 %

Source of chirality : Chemical synthesis from the corresponding D- or L- α - amino acid
Absolute configuration : R or S

Enantioseparation of 5-Monosubstituted Hydantoins by Capillary Gas Chromatography - Investigation of Chemical and Enzymatic Racemization

Tetrahedron: Asymmetry 1993, 4, 1129

H. Lickefett¹, K. Krohn², W.A. König³, B. Gehrcke³ and C. Syldatk¹



C₆H₁₀N₂O₂
5-Isopropyl-imidazolidin-2,4-dione

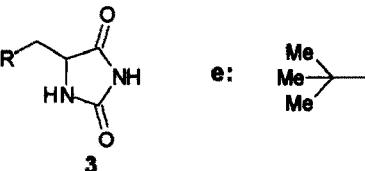
E. e > 98 %

Source of chirality : Chemical synthesis from the corresponding D- or L- α - amino acid
Absolute configuration : R or S

Enantioseparation of 5-Monosubstituted Hydantoins by Capillary Gas Chromatography - Investigation of Chemical and Enzymatic Racemization

Tetrahedron: Asymmetry 1993, 4, 1129

H. Lickefett¹, K. Krohn², W.A. König³, B. Gehrcke³ and C. Syldatk¹



C₈H₁₄N₂O₂
5-tert-butyl-imidazolidin-2,4-dione

E. e > 98 %

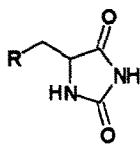
[α]_D²⁰ +63.9 (c = 0.27, ethanol)

Source of chirality : Chemical synthesis from the corresponding D- or L- α - amino acid
Absolute configuration : R or S

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f:



C₉H₁₄N₂O₂

5-Cyclohexyl-methyl-imidazolidin-2,4-dione

E. e > 98 %

[α]_D²⁰ +62.4 (c = 1.27, ethanol)

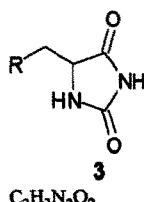
Source of chirality : Chemical synthesis from the corresponding D- or L- α - amino acid

Absolute configuration : R or S

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Tetrahedron: Asymmetry 1993, 4, 1129



g:



C₉H₁₄N₂O₂

5-Phenyl-imidazolidin-2,4-dione

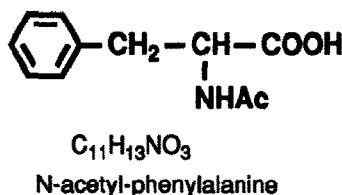
E. e > 98 %

Source of chirality : Chemical synthesis from the corresponding D- or L- α - amino acid

Absolute configuration : R or S

R. Chênevert, R. BelRhïd, M. Létourneau, R. Gagnon, L. D'Astous.

Tetrahedron: Asymmetry 1993, 4, 1137



C₁₁H₁₃NO₃

N-acetyl-phenylalanine

E.e. > 95% (¹H NMR of (S)-naphthylethylamide)

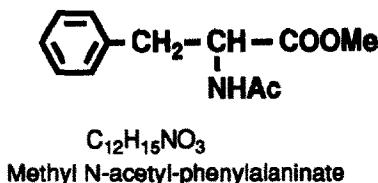
[α]_D²³ = -51.0 (c 4, EtOH)

Source of chirality : enzymatic hydrolysis

Absolute configuration : D

R. Chênevert, R. BelRhïd, M. Létourneau, R. Gagnon, L. D'Astous.

Tetrahedron: Asymmetry 1993, 4, 1137



C₁₂H₁₅NO₃

Methyl N-acetyl-phenylalaninate

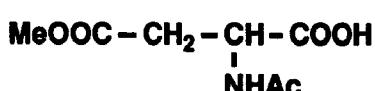
E.e > 95% (optical rotation)

[α]_D²³ = +19.3 (c 3, MeOH)

Source of chirality : enzymatic hydrolysis

Absolute configuration : L

R. Chênevert, R. BelRhïid, M. Létourneau, R. Gagnon, L. D'Astous.



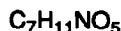
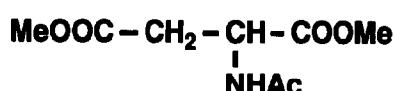
B-methyl N-acetyl-aspartate

E.e > 95% (^1H NMR of (S)-1-naphthylethylamide)

$[\alpha]_D^{23} = -8.3$ (c 3, EtOH)

Source of chirality : enzymatic hydrolysis

Absolute configuration : D



Dimethyl N-acetyl-aspartate

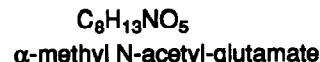
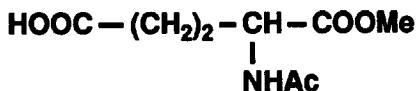
E.e > 95% (optical rotation)

$[\alpha]_D^{23} = -21.0$ (c 1, CHCl_3)

Source of chirality : enzymatic hydrolysis

Absolute configuration : L

R. Chênevert, R. BelRhïid, M. Létourneau, R. Gagnon, L. D'Astous.



α -methyl N-acetyl-glutamate

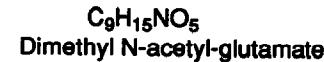
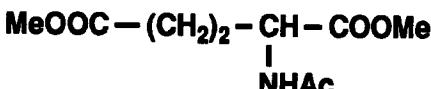
E.e > 95% (^1H NMR of (S)-1-naphthylethylamide)

$[\alpha]_D^{23} = +23.0$ (c 4, MeOH)

Source of chirality : enzymatic hydrolysis

Absolute configuration : D

R. Chênevert, R. BelRhïid, M. Létourneau, R. Gagnon, L. D'Astous.



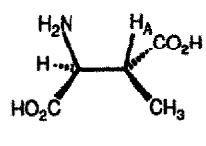
Dimethyl N-acetyl-glutamate

E.e > 95% (optical rotation)

$[\alpha]_D^{23} = +12.2$ (c 3, MeOH)

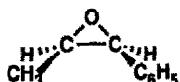
Source of chirality : enzymatic hydrolysis

Absolute configuration : L



3-Methylaspartic acid

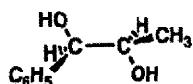
D.e. 95% (asymmetric alkylation) or
98% (enzymic amination then resolution)
100% (2S)-configuration
 $\text{H}_A = \text{H}; [\alpha]_D^{20} +36.3$ (c 1.0, 5 M HCl)
 $\text{H}_A = {}^2\text{H}; [\alpha]_D^{20} +30.5$ (c 1.0, 5 M HCl)
 Source of chirality: asymm. synth. or enzyme
 Absolute configuration 2S,3R

 $\text{C}_9\text{H}_{10}\text{O}$

cis-1-Phenylpropene oxide

E.e. >98% [by glc: Chiraldex G-TA]

Source of chirality: enzymatic hydrolysis of the racemate
 Absolute configuration (1S,2R)

 $\text{C}_9\text{H}_{12}\text{O}_2$

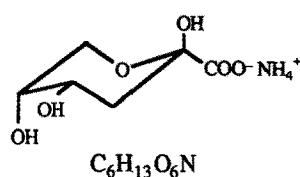
threo-1-Phenylpropane-1,2-diol

E.e. >98% [by glc: Chiraldex G-TA]

Source of chirality: enzymatic hydrolysis of (\pm)-cis-1-phenylpropene oxide
 Absolute configuration (1R,2R)

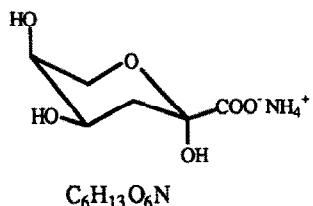
 $\text{C}_8\text{H}_{16}\text{O}$
2-Methyl-2-pentyloxiran

E.e. = 72% [by chiral GC]
 $[\alpha]_D^{20} = -6.82$ (c 3.43, CHCl_3)
 Source of chirality: enzymatic resolution.
 Absolute configuration: (*R*) by comparison with independently synthesized material.

D.e. = 92% (by ^1H and ^{13}C NMR) $[\alpha]_D^{20} = -26$ (c, 2, water)

Source of chirality: microbiological aldolisation

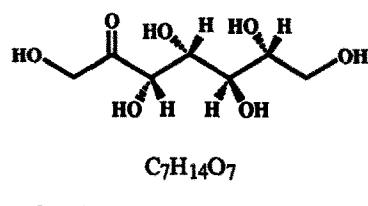
Absolute configuration 4S, 5R

Diastereomeric mixture (by ^1H and ^{13}C NMR):

60:40 4R, 5S:4S, 5S

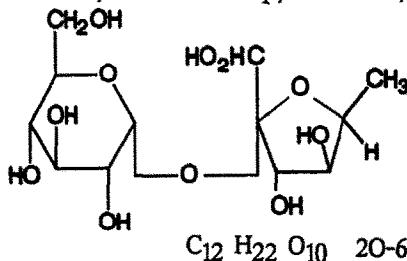
 $[\alpha]_D^{20} = +7.9$ (c, 1.25, water)

Source of chirality: microbiological aldolisation

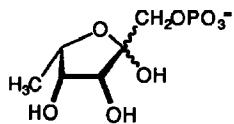
 $[\alpha]_D^{25} = +8$ (c = 0.03, H_2O)

Source of chirality : Transketolase

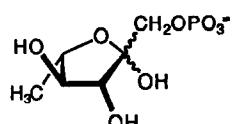
Absolute configuration : 3S, 4R, 5R, 6R

(assigned on the basis of $[\alpha]_D^{25}$)

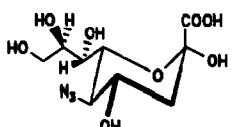
Source of chirality: natural and enzymatic synthesis

 $C_6H_{11}O_8P \times 2 (C_6H_{14}N)$ 6-Deoxy-L-lyxo-hexulose 1-phosphate,
bis(cyclohexylammonium) saltE.e. = 100%
 $[\alpha]_D^{18} = +0.5$ (c 2, H₂O)

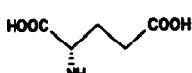
Source of chirality: natural (from L-fucose) and enzymatic synthesis (aldol addition)

Absolute configuration 3R,4R,5S
by relation to natural L-fucose $C_6H_{11}O_8P \times 2 (C_6H_{14}N)$ 6-Deoxy-L-arabino-hexulose 1-phosphate,
bis(cyclohexylammonium) saltE.e. = 100%
 $[\alpha]_D^{18} = +2$ (c 1, H₂O)

Source of chirality: natural (from L-rhamnose) and enzymatic synthesis (aldol addition)

Absolute configuration 3R,4S,5S
by relation to natural L-rhamnose $C_9H_{15}N_3O_5$ 5-Azido-3,5-dideoxy-D-glycero-D-galacto-nonulonic acid
5-Azido-neuraminic acid $[\alpha]_D^{20} = -62.2$ (c = 0.66, H₂O)

Source of chirality: natural and enzymatic asymmetric aldol condensation

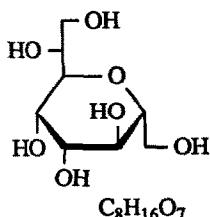
 $C_5H_9NO_4$ ¹⁵N-L-Glutamic acid

E.e. > 99.5 % [by HPLC]

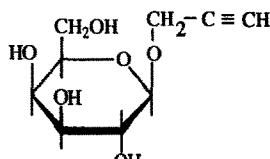
Source of chirality: enzymatic asymmetric reduction
(reductive amination)

Absolute configuration: 2S

¹⁵N content 98 %

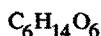
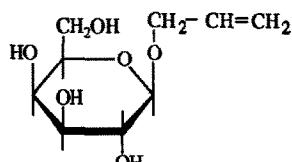
*L-allo-D-erythro-3,7-anhydro-octitol*

e.d. = about 100%
 $[\alpha]_D = +6.6$ (c 1, MeOH)
 Source of chirality: D-ribose 5-phosphate
 Absolute configuration: 2*R*,3*S*,4*S*,5*S*,6*R*,7*R*
 (assigned by 1H NMR and reaction mechanism)



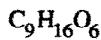
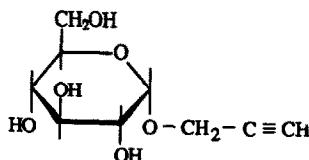
$[\alpha]_D^{25} = -39.8$ (c1.8, H₂O)

Source of chirality: enzymatic synthesis from lactose

Propargyl- β -D-Galactopyranoside

$[\alpha]_D^{25} = -11.2$ (c2, H₂O)

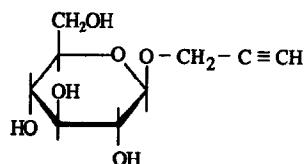
Source of chirality: enzymatic synthesis from lactose

Allyl- β -D-Galactopyranoside

$[\alpha]_D^{25} = +95.9$ (c1.8, H₂O)

Source of chirality: enzymatic synthesis from maltose

Propargyl- α -D-glucopyranoside


 $[\alpha]_D^{25} = -54.5 \text{ (c } 1.8, \text{ H}_2\text{O)}$

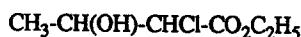
Source of chirality: enzymatic synthesis from cellobiose

Propargyl- β -D-Glucopyranoside

M. Hamdani, B. De Jeso, H. Deleuze,

A. Saux and B. Maillard.

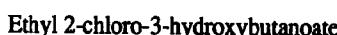
Sources of chirality :



- Reduction of the corresponding ketoester by baker's yeast
d.e. = 96% (2R,3S) e.e. = 96%



- D-Threonine
d.e. > 98% (2R,3S) e.e. > 96%

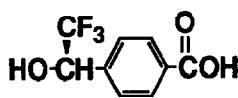

 $[\alpha]_D^{25} = +12.4 \text{ (c } 1, \text{ CHCl}_3)$

T. Fujisawa,* K. Ichikawa, and M. Shimizu

ee = >99% (after recrystallization from toluene) [determined by GLC analysis of the corresponding MTPA ester]

$[\alpha]_D^{23} -28.1 \text{ (c } 0.08, \text{ MeOH)}$

Source of chirality: Bakers' yeast reduction



Absolute configuration: R (assigned by comparison with the authentic sample prepared from the known (R)-p-(2,2,2-trifluoro-1-hydroxyethyl)bromobenzene)

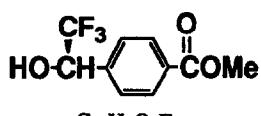
(R)-p-(2,2,2-Trifluoro-1-hydroxyethyl)benzoic Acid

T. Fujisawa,* K. Ichikawa, and M. Shimizu

ee = >99% (after recrystallization from n-hexane) [determined by GLC analysis of the corresponding MTPA ester]

$[\alpha]_D^{23} -28.3 \text{ (c } 0.12, \text{ MeOH)}$

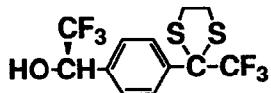
Source of chirality: Bakers' yeast reduction



Absolute configuration: R (assigned by comparison with the authentic sample prepared from the known (R)-p-(2,2,2-trifluoro-1-hydroxyethyl)bromobenzene)

Methyl (R)-p-(2,2,2-Trifluoro-1-hydroxyethyl)benzoate

T. Fujisawa,* K. Ichikawa, and M. Shimizu

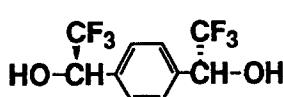
ee = >99% (after recrystallization from toluene) [determined by HPLC analysis
of the corresponding MTPA ester] $C_{12}H_{10}OS_2F_6$ $[\alpha]_D^{23} -21.0$ (c 0.20, MeOH)

Source of chirality: Bakers' yeast reduction

Absolute configuration: *R* (assigned by comparison with the authentic sample prepared from the known (*R*)-*p*-(2,2,2-trifluoro-1-hydroxyethyl)bromobenzene) (*R*)-4-(2,2,2-Trifluoro-1-hydroxyethyl)-1-(2-trifluoromethyl-1,3-dithiolan-2-yl)benzene

T. Fujisawa,* K. Ichikawa, and M. Shimizu

ee = >99% (after recrystallization from carbon tetrachloride) [determined by

 $C_{10}H_8O_2F_6$

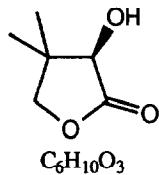
GLC analysis of the corresponding MTPA ester

 $[\alpha]_D^{23} -48.0$ (c 0.10, MeOH)

Source of chirality: Bakers' yeast reduction

Absolute configuration: *R,R* (assigned by comparison with the authentic sample prepared from the known (*R*)-*p*-(2,2,2-trifluoro-1-hydroxyethyl)bromobenzene) (*R,R*)-*p*-Bis(2,2,2-trifluoro-1-hydroxyethyl)benzene

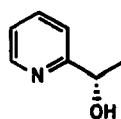
K. Nakamura, S. Kondo, Y. Kawai, and A. Ohno

 $C_6H_{10}O_3$ E.e. = 93% [by chiral HPLC analysis of corresponding
3,5-dinitrobenzoyl ester]

Source of chirality : Microbial reduction

Absolute configuration : *R*(R)-(-)-Pantolactone
(R)-Dihydro-3-hydroxy-4,4-dimethyl-2(3*H*)-furanone

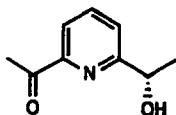
D. Bailey, D. O'Hagan, U. Dyer and R. B. Lamont

E.e. = >95 [by n m r of acetate with Eu(hfc)3]
 $[\alpha]_D^{20} = -29.14$ (c4.94, CHCl3)

Source of chirality: Bakers' yeast reduction

Absolute configuration (S)

 C_7H_9NO
2-alpha-ethanolpyridine



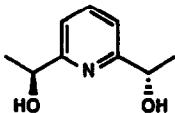
E.e. = 99.8 [chiral HPLC analysis]
 $[\alpha]_D^{20} = -7.5$ (c1.5, CHCl₃)

Source of chirality: Bakers' yeast reduction

Absolute configuration (S)

C₉H₁₁NO₂

2-Acetyl-6-alpha-ethanolpyridine



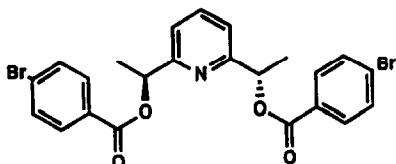
E.e. = 99.97 [chiral HPLC analysis]
 $[\alpha]_D^{20} = -26.84$ (c2.98, CHCl₃)

Source of chirality: Bakers' yeast reduction

Absolute configuration (S,S)
 (assigned by X-ray of di-p-bromobenzoate)

C₉H₁₃NO₂

2,6-di-alpha-ethanolpyridine



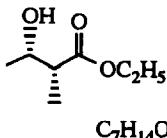
E.e. = 99.97 [chiral HPLC analysis]
 $[\alpha]_D^{20} = +70.18$ (c1.71, CHCl₃)
 mp 154-154.5°C

Source of chirality: Bakers' yeast reduction

Absolute configuration (S,S)
 (assigned by X-ray analysis)

C₂₃H₁₉Br₂NO₄

2,6-diethylpyridine di-alpha,alpha,-p-bromobenzoate



Ethyl 2-methyl-3-hydroxybutanoate

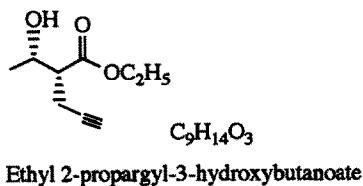
D.e. = 82% [by GC]

E.e. = 98% [by HPLC analysis of the Mosher's ester]

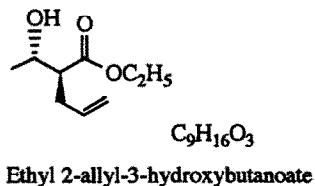
$[\alpha]_D^{23} = +6.15$ (c 2.42, CHCl₃)

Source of chirality: yeast reduction

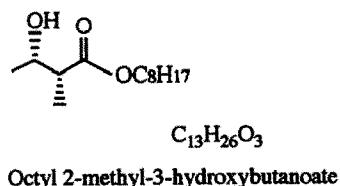
Absolute configuration: 2R,3S



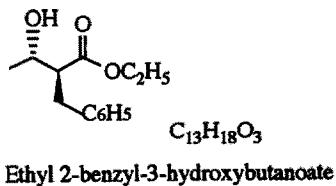
D.e. = 52% [by GC]
 E.e. = 98% [by HPLC analysis of the Mosher's ester]
 $[\alpha]_D^{23} = +23.13$ (c 2.35, CHCl_3)
 Source of chirality: yeast reduction
 Absolute configuration: 2R,3S



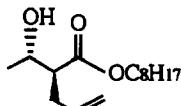
D.e. = 54% [by GC]
 E.e. = 98% [by HPLC analysis of the Mosher's ester]
 $[\alpha]_D^{23} = +12.61$ (c 1.83, CHCl_3)
 Source of chirality: yeast reduction
 Absolute configuration: 2S,3S



D.e. = 92% [by GC]
 E.e. = 98% [by HPLC analysis of the Mosher's ester]
 $[\alpha]_D^{23} = +3.30$ (c 3.18, CHCl_3)
 Source of chirality: yeast reduction
 Absolute configuration: 2R,3S



D.e. = 40% [by GC]
 E.e. = 98% [by HPLC analysis of the Mosher's ester]
 $[\alpha]_D^{23} = -9.03$ (c 2.9, CHCl_3)
 Source of chirality: yeast reduction
 Absolute configuration: 2S,3S

C₁₅H₂₈O₃

Octyl 2-allyl-3-hydroxybutanoate

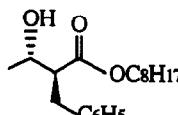
D.e. = 20% [by GC]

E.e. = 98% [by HPLC analysis of the Mosher's ester]

[α]_D²³ = +5.10 (c 1.88, CHCl₃)

Source of chirality: yeast reduction

Absolute configuration: 2S,3S

C₁₉H₃₀O₃

Octyl 2-benzyl-3-hydroxybutanoate

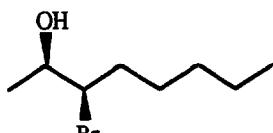
D.e. = 88% [by GC]

E.e. = 98% [by HPLC analysis of the Mosher's ester]

[α]_D²³ = -24.31 (c 0.54, CHCl₃)

Source of chirality: yeast reduction

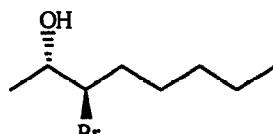
Absolute configuration: 2S,3S

C₈H₁₇BrO
(2R,3R)-3-bromo-2-octanolE.e. > 98 % (by GC analysis of esters obtained with
(+)-(S)-O-acetyl lactic acid chloride)[α]_D²⁵ = + 38 (c = 0.02, CHCl₃)

Source of chirality : Microbiological reduction

Absolute configuration : 2R, 3R

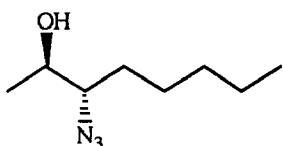
(assigned by chemical correlation)

C₈H₁₇BrO
(2S,3R)-3-bromo-2-octanolE.e. > 98 % (by GC analysis of esters obtained with
(+)-(S)-O-acetyl lactic acid chloride)[α]_D²⁵ = + 40 (c = 0.02, CHCl₃)

Source of chirality : Microbiological reduction

Absolute configuration : 2S, 3R

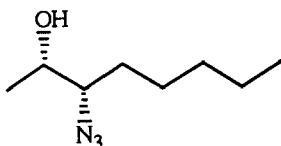
(assigned by chemical correlation)



C₈H₁₇N₃O
(2R,3S)-3-azido-2-octanol

E.e. > 98 % (by GC analysis with chiral column : Lipodex E)
 $[\alpha]_D^{25} = -8$ (*c* = 0.01, CHCl₃)

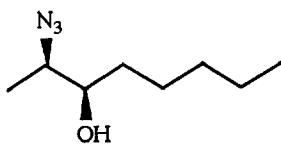
Source of chirality : Microbiological reduction
 Absolute configuration : 2R, 3S
 (assigned by chemical correlation)



C₈H₁₇N₃O
(2S,3S)-3-azido-2-octanol

E.e. > 98 % (by GC analysis with chiral column : Lipodex E)
 $[\alpha]_D^{25} = +21$ (*c* = 0.03, CHCl₃)

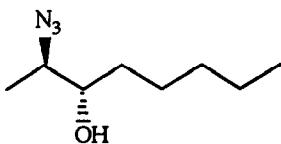
Source of chirality : Microbiological reduction
 Absolute configuration : 2S, 3S
 (assigned by chemical correlation)



C₈H₁₇N₃O
(2R,3R)-2-azido-3-octanol

E.e. > 98 % (by GC analysis with chiral column : Lipodex E)
 $[\alpha]_D^{25} = -50$ (*c* = 0.03, CHCl₃)

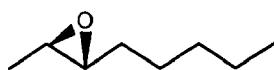
Source of chirality : from a precursor obtained by
 microbiological reduction
 Absolute configuration : 2R, 3R
 (assigned based on the reaction mechanism)



C₈H₁₇N₃O
(2R,3S)-2-azido-3-octanol

E.e. > 98 % (by GC analysis with chiral column : Lipodex E)
 $[\alpha]_D^{25} = -51$ (*c* = 0.02, CHCl₃)

Source of chirality : from a precursor obtained by
 microbiological reduction
 Absolute configuration : 2R, 3S
 (assigned based on the reaction mechanism)

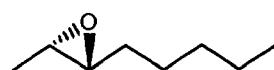


C₈H₁₆O
(2R,3S)-2,3-epoxyoctane

E.e. > 98 % (by GC analysis with chiral column : Lipodex E)
[α]_D²⁵ = - 8 (c = 0.04, Pentane)

Source of chirality : from a precursor obtained by
microbiological reduction

Absolute configuration : 2R, 3S
(assigned based on the reaction mechanism)

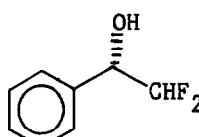


C₈H₁₆O
(2S,3S)-2,3-epoxyoctane

E.e. > 98 % (by GC analysis with chiral column : Lipodex E)
[α]_D²⁵ = + 5 (c = 0.03, Pentane)

Source of chirality : from a precursor obtained by
microbiological reduction

Absolute configuration : 2S, 3S
(assigned based on the reaction mechanism)



C₈H₈F₂O

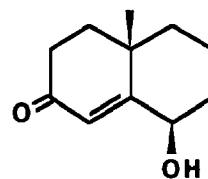
α-(Difluoromethyl)benzyl alcohol

E.e. = 97 % (by HPLC)

[α]_D²⁷ = + 19 (c = 2.8, CH₂Cl₂)

Source of chirality : enzymic reduction

Absolute configuration: S



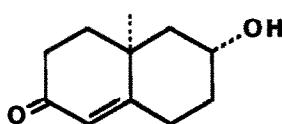
C₁₁H₁₆O₂
4a-methyl-8-hydroxy-4,4a,5,6,7,8-hexahydro-2(3H)-naphthalenone

E.e. ≥ 95% (from e.e. of the precursor)

[α]_D²¹ = + 96.5 (c 1.025, CHCl₃)

Source of chirality: microbial hydroxylation of the corresponding (S)-naphthalenone

Absolute configuration: 4aS,8R (relative configuration assigned by NMR)



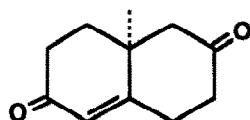
C₁₀H₁₅O₂
4a-methyl-6-hydroxy-4,4a,5,6,7,8-hexahydro-2(3H)-naphthalenone

E.e. ≥ 95% (from e.e. of the precursor)

[α]_D²¹ = -210 (c 1.025, CHCl₃)

Source of chirality: microbial hydroxylation of the corresponding (R)-naphthalenone

Absolute configuration: 4aS,6R (relative configuration assigned by NMR)



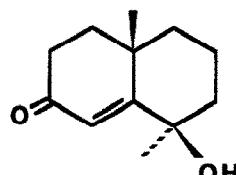
C₁₁H₁₄O₂
4a-methyl-4,4a,7,8-tetrahydro-naphthalene-2(3H), 6(5H)-dione

E.e. ≥ 95% (from e.e. of the precursor)

[α]_D²¹ = -115 (c 0.3, CHCl₃)

Source of chirality: microbial hydroxylation of the corresponding (R)-naphthalenone

Absolute configuration: 4aS



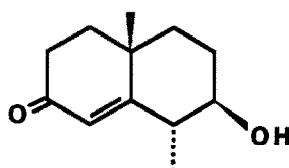
C₁₂H₁₈O₂
4a,8-dimethyl-8-hydroxy-4,4a,5,6,7,8-hexahydro-2(3H)-naphthalenone

E.e. ≥ 95% (from e.e. of the precursor)

[α]_D²¹ = + 71 (c 0.21, CHCl₃)

Source of chirality: microbial hydroxylation of the corresponding (4aS,8S)-naphthalenone

Absolute configuration: 4aS,8R (relative configuration assigned by NMR)



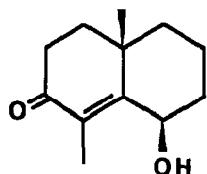
C₁₂H₁₈O₂
4a,8-dimethyl-7-hydroxy-4,4a,5,6,7,8-hexahydro-2(3H)-naphthalenone

E.e. ≥ 95% (from e.e. of the precursor)

[α]_D²¹ = + 62 (c 0.47, CHCl₃)

Source of chirality: microbial hydroxylation of the corresponding (4aS,8S)-naphthalenone

Absolute configuration: 4aS,7R (relative configuration assigned by NMR)



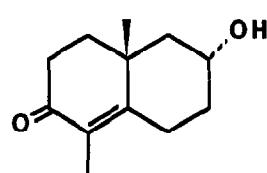
C₁₂H₁₆O₂
1,4a-dimethyl-8-hydroxy-4,4a,5,6,7,8-
hexahydro-2(3H)-naphthalenone

E.e. ≥ 90% (from e.e. of the precursor)

[α]_D²¹ = +40 (c 3.7, CHCl₃)

Source of chirality: microbial hydroxylation of the
corresponding (S)-naphthalenone

Absolute configuration: 4aS,8R (relative configuration
assigned by NMR)



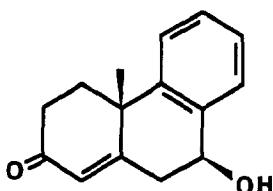
C₁₂H₁₆O₂
1,4a-dimethyl-6-hydroxy-4,4a,5,6,7,8-
hexahydro-2(3H)-naphthalenone

E.e. ≥ 90% (from e.e. of the precursor)

[α]_D²¹ = +164 (c 1.34, CHCl₃)

Source of chirality: microbial hydroxylation of the
corresponding (S)-naphthalenone

Absolute configuration: 4aR,6R (relative configuration assigned
by NMR)



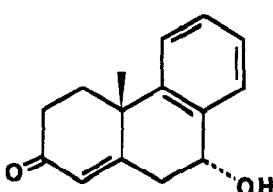
C₁₅H₁₆O₂
4a-methyl-9-hydroxy-4,4a,9,10-
tetrahydro-2(3H)-phenanthrenone

E.e. ≥ 90% (from e.e. of the precursor)

[α]_D²¹ = +208 (c 1.85, CHCl₃)

Source of chirality: microbial hydroxylation of the
corresponding (S)-phenanthrenone

Absolute configuration: 4aS,6S (relative configuration
assigned by NMR)



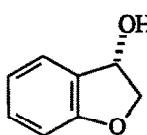
C₁₅H₁₆O₂
4a-methyl-9-hydroxy-4,4a,9,10-
tetrahydro-2(3H)-phenanthrenone

E.e. ≥ 90% (from e.e. of the precursor)

[α]_D²¹ = +105 (c 0.6, CHCl₃)

Source of chirality: microbial hydroxylation of the
corresponding (S)-phenanthrenone

Absolute configuration: 4aS,6R (relative configuration
assigned by NMR)



C₈H₈O₂
3-Hydroxy-2,3-dihydrobenzofuran

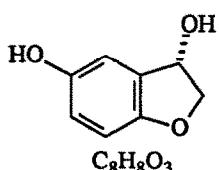
E.e.= >98% [by CSP-HPLC analysis]

[α]D = +67 (c 0.63 , CHCl₃)

Source of chirality : Resolution of the camphanate esters .

Absolute configuration : 3S

(Assigned by X-ray structure analysis of the camphanate ester)



3,5-Dihydroxy-2,3-dihydrobenzofuran

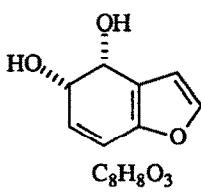
E.e.= > 98% [by CSP-HPLC analysis and by ¹H-NMR analysis of di-MTPA esters].

[α]D = +22.5 , (c 0.67 ,MeOH)

Source of chirality : Biotransformation of enantiopure (+)-3-hydroxy-2,3-dihydrobenzofuran.

Absolute configuration : 3S

[Derived from (3S)-3-hydroxy-2,3-dihydrobenzofuran].



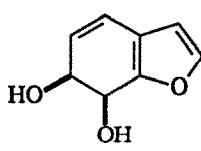
C₈H₈O₃
cis-4,5-Dihydroxy-4,5-dihydrobenzofuran

E.e.= >98% [by ¹H-NMR analysis of di-MTPA esters].

Source of chirality : Biotransformation of enantiopure (+)-3-hydroxy-2,3-dihydrobenzofuran.

Absolute configuration : 4R,5S

[Assigned by stereochemical correlation]



C₈H₈O₃
cis-6,7-Dihydroxy-6,7-dihydrobenzofuran

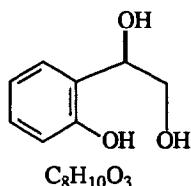
E.e.= >98% [by ¹H-NMR analysis of di-MTPA esters].

[α]D = -35 (c 0.96, MeOH).

Source of chirality : Biotransformation of benzofuran

Absolute configuration : 6S,7S

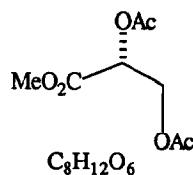
[Assigned by stereochemical correlation]

E.e.= 51% (by [α]_D comparison)[α]_D = - 24 (c 0.76, MeOH)

Source of chirality : Biotransformation of benzofuran.

1,2-Dihydroxy-1-(2'-hydroxyphenyl)ethane

Absolute configuration : 1R

[Assigned by stereochemical correlation with (2R)-methyl
(2,3-diacetoxy)propanoate].

E.e.= >98% (by synthesis from enantiopure(R)-glyceric acid.)

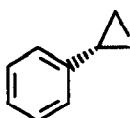
[α]_D = +15.8 (c 2.0 ,CHCl₃)

Source of chirality : Synthesis from enantiopure(R)- glyceric acid.

Absolute configuration : 2R

[Assigned by stereochemical correlation with(R)-glyceric acid].

Methyl (2,3-diacetoxy)propanoate

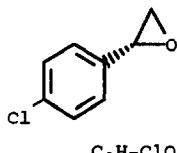


E.e.=49% (by chiral HPLC with Chiralcel OB column)

Source of chirality: Chloroperoxidase

Absolute configuration: R

Styrene oxide

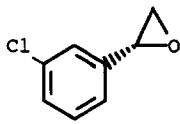


E.e.=66% (by chiral HPLC with Chiralcel OB column)

Source of chirality: Chloroperoxidase

Absolute configuration: R

4-Chlorostyrene oxide



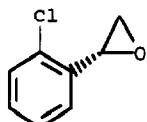
E.e.=62% (by chiral HPLC with Chiralcel OB column)

Source of chirality: Chloroperoxidase

Absolute configuration: R



3-Chlorostyrene oxide



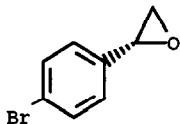
E.e.=64% (by chiral HPLC with Chiralcel OB column)

Source of chirality: Chloroperoxidase

Absolute configuration: R



2-Chlorostyrene oxide



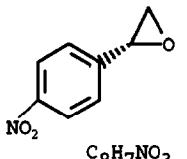
E.e.=68% (by chiral HPLC with Chiralcel OB column)

Source of chirality: Chloroperoxidase

Absolute configuration: R



4-Bromostyrene oxide



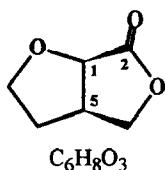
E.e.=28% (by chiral HPLC with Chiralcel OB column)

Source of chirality: Chloroperoxidase

Absolute configuration: R



4-Nitrostyrene oxide



3,8-dioxabicyclo[3.3.0]octan-2-one.

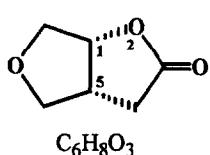
E.e. > 98 % (by chiral GC)
 $[\alpha]_D^{25} - 103.2$ ($c = 0.5$, CHCl_3)

Source of chirality : enzymatic

Baeyer-Villiger oxidation .

Absolute configuration : 1R,5S

(assigned by circular dichroism measurement).



2,7-dioxabicyclo[3.3.0]octan-3-one.

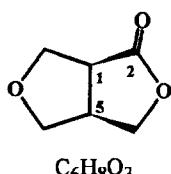
E.e. 97 % (by chiral GC)
 $[\alpha]_D^{25} - 37$ ($c = 0.5$, CHCl_3)

Source of chirality : enzymatic

Baeyer-Villiger oxidation

Absolute configuration : 1R,5R

(assigned by circular dichroism measurement).



3,7-dioxabicyclo[3.3.0]octan-2-one.

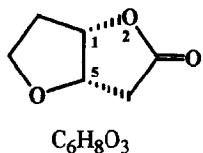
E.e.> 98 % (by chiral GC)
 $[\alpha]_D^{25} - 101.2$ ($c = 0.5$, CHCl_3)

Source of chirality : enzymatic

Baeyer-Villiger oxidation .

Absolute configuration : 1S,5R

(assigned by circular dichroism measurement).



2,6-dioxabicyclo[3.3.0]octan-3-one.

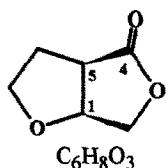
E.e.> 98 % (by chiral GC)
 $[\alpha]_D^{25} - 67.3$ ($c = 0.639$, CHCl_3)

Source of chirality : enzymatic

Baeyer-Villiger oxidation .

Absolute configuration : 1S,5S

(assigned by circular dichroism measurement)



3,8-dioxabicyclo[3.3.0]octan-4-one.

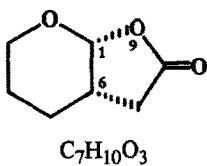
E.e. > 98 % (by chiral GC)

 $[\alpha]_D^{25} - 113.3$ ($c = 0.6$, CHCl_3)*Source of chirality* : enzymatic

Baeyer-Villiger oxidation .

Absolute configuration : 1S,5R

(assigned by circular dichroism measurement).



2,9-dioxabicyclo[4.3.0]nonan-8-one.

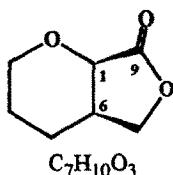
E.e. 70 % (by chiral GC)

 $[\alpha]_D^{25} - 4$ ($c = 0.55$, CHCl_3)*Source of chirality* : enzymatic

Baeyer-Villiger oxidation .

Absolute configuration : 1R,6S

(assigned by circular dichroism measurement).



2,8-dioxabicyclo[4.3.0]nonan-9-one.

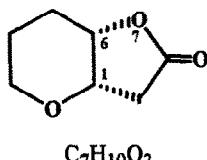
E.e. > 98 % (by chiral GC)

 $[\alpha]_D^{25} - 105.1$ ($c = 0.69$, CHCl_3)*Source of chirality* : enzymatic

Baeyer-Villiger oxidation .

Absolute configuration : 1R,6S

(assigned by circular dichroism measurement).



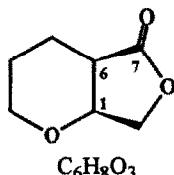
2,7-dioxabicyclo[3.3.0]octan-8-one.

E.e. 33 % (by chiral GC)

 $[\alpha]_D^{25} - 24$ ($c = 0.5$, CHCl_3)*Source of chirality* : enzymatic

Baeyer-Villiger oxidation .

Absolute configuration : 1S,6R



E.e. > 98 % (by chiral GC)

$[\alpha]_D^{25} - 26.5$ ($c = 0.577$, $CHCl_3$)

Source of chirality : enzymatic

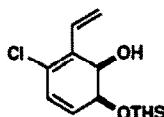
Baeyer-Villiger oxidation .

Absolute configuration : 1S,6R

(assigned by circular dichroism measurement).

2,8-dioxabicyclo[4.3.0]nonan-7-one.

T. Hudlicky, E. E. Boros, C. H. Boros, Department of Chemistry
Virginia Polytechnic Institute and State University, Blacksburg, VA 24061-0212



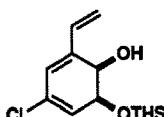
$[\alpha]_D = +101.1$ ($c 1.12$, $CHCl_3$)

E.e. >98%

Obtained from *Pp*-39D oxidation of *o*-chlorostyrene and protection.

Absolute configuration established by convergent synthesis.

T. Hudlicky, E. E. Boros, C. H. Boros, Department of Chemistry
Virginia Polytechnic Institute and State University, Blacksburg, VA 24061-0212



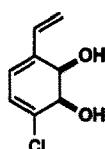
$[\alpha]_D = +82.9$ ($c 0.42$, $CHCl_3$)

E.e. 54%

Obtained from *Pp*-39D oxidation of *m*-chlorostyrene and protection.

Absolute configuration established by convergent synthesis.

T. Hudlicky, E. E. Boros, C. H. Boros, Department of Chemistry
Virginia Polytechnic Institute and State University, Blacksburg, VA 24061-0212



$[\alpha]_D = +12.4$ ($c 0.064$, $CHCl_3$)

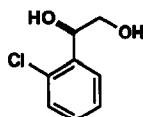
E.e. 15%

Obtained from *Pp*-39D oxidation of *p*-chlorostyrene.

Absolute configuration established by convergent synthesis.

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Tetrahedron: Asymmetry 1993, 4, 1365

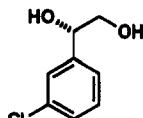


$[\alpha_D] = -47.2$ (c 1.9, EtOH)
E.e. 73%

Obtained from *Pp*-39D oxidation of *o*-chlorostyrene.
Absolute configuration established by convergent synthesis.

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Tetrahedron: Asymmetry 1993, 4, 1365

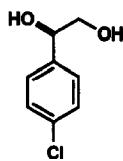


$[\alpha_D] = +24.05$ (c 1.24, EtOH)
E.e. 95%

Obtained from *Pp*-39D oxidation of *m*-chlorostyrene.
Absolute configuration established by convergent synthesis.

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Virginia Polytechnic Institute and State University, Blacksburg, VA 24061-0212

Tetrahedron: Asymmetry 1993, 4, 1365

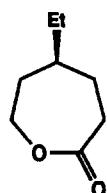


$[\alpha_D] = -27.60$ (c 0.96, EtOH)
E.e. 79%

Obtained from *Pp*-39D oxidation of *p*-chlorostyrene.
Absolute configuration established by convergent synthesis.

Michael J. Taschner*, Donald J. Black, and Quin-Zene Chen

Tetrahedron: Asymmetry 1993, 4, 1387



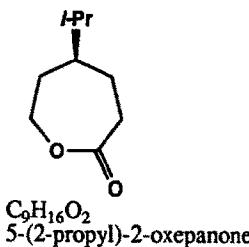
C₈H₁₄O₂
5-ethyl-2-oxepanone

E.e. > 98% (by ¹H-NMR of a MTPA ester derivative)

$[\alpha]_D = -38$ (c 5.55, CHCl₃)

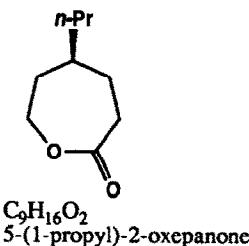
Source of chirality: Enzymatic Baeyer-Villiger oxidation

Absolute Configuration: 5*S* (assignment tentative)

E.e. > 98% (by $^1\text{H-NMR}$ of a MTPA ester derivative) $[\alpha]_D = -40$ (c 0.44, CHCl_3)

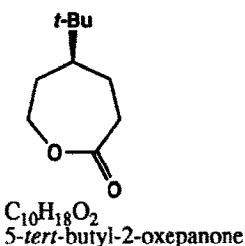
Source of chirality: Enzymatic Baeyer-Villiger oxidation

Absolute Configuration: 5S (assignment tentative)

E.e. > 98% (by $^1\text{H-NMR}$ of a MTPA ester derivative) $[\alpha]_D = -38$ (c 6.41, CHCl_3)

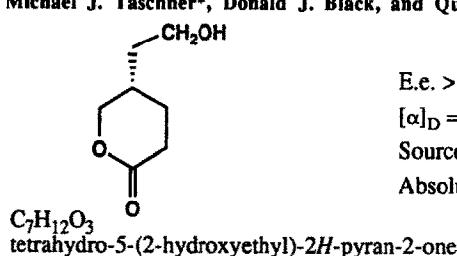
Source of chirality: Enzymatic Baeyer-Villiger oxidation

Absolute Configuration: 5S (assignment tentative)

E.e. > 98% (by $^1\text{H-NMR}$ of a MTPA ester derivative) $[\alpha]_D = -34.9$ (c 0.78, CHCl_3)

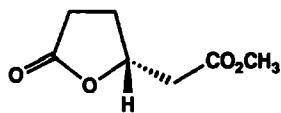
Source of chirality: Enzymatic Baeyer-Villiger oxidation

Absolute Configuration: 5S (assignment via chemical correlation)

E.e. > 98% (by $^1\text{H-NMR}$ of a MTPA ester derivative) $[\alpha]_D = -6.2$ (c 5.66, CHCl_3)

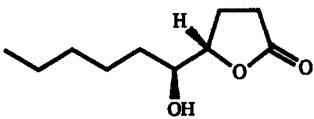
Source of chirality: Enzymatic Baeyer-Villiger oxidation

Absolute Configuration: 5S (assignment tentative)



E.e. = 9.6% (by $^1\text{H-NMR}$ of a MTPA ester derivative)
 $[\alpha]_D = -3.65$ (c 0.99, EtOH)
 Source of chirality: Enzymatic Baeyer-Villiger oxidation
 Absolute Configuration: 2*R* (assignment via chemical correlation)

$\text{C}_7\text{H}_{10}\text{O}_4$
 tetrahydro-5-oxo-2-furanacetic acid methyl ester

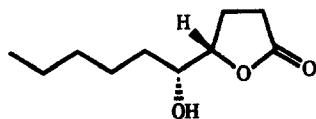


E.e. = 92.4 % [by GC after conversion into the carbamate derivative with (R)-(+)-1-Phenylethylisocyanate]

$[\alpha]_D = +26.2$ (c 1.74, CHCl_3)

Source of chirality: (S)-glutamic acid

Absolute configuration: 4*S*, 5*S*



E.e. = not determined
 $[\alpha]_D = +13.3$ (c 1.35, CHCl_3)

Source of chirality: (S)-glutamic acid

Absolute configuration: 4*S*, 5*R*